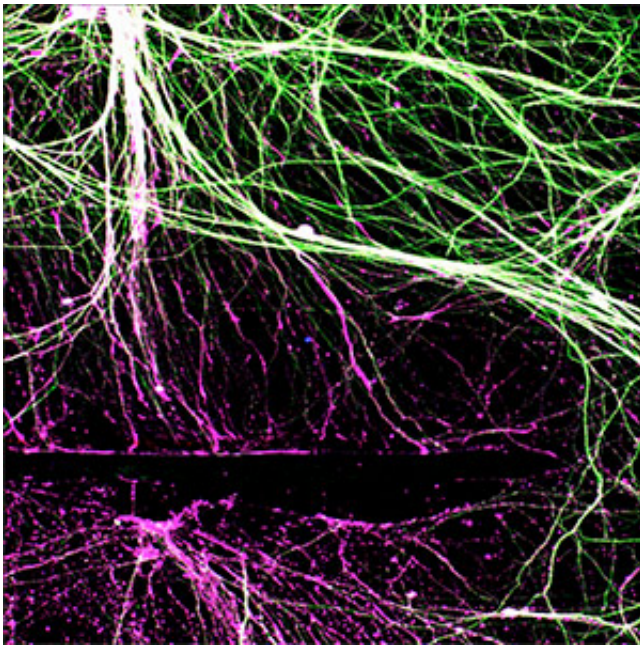


Master gene orchestrates regeneration of damaged peripheral nerves

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To study how peripheral neurons regrow axons — the branches of nerve cells that transmit nerve signals — researchers at Washington University School of Medicine in St. Louis grow them in a dish, cut them (as shown) and observe how they regrow. The researchers, led by Valeria Cavalli, PhD, have identified a master gene that enables repair of these branches when they are cut. Credit: Cavalli Laboratory

One of the big challenges with spinal cord injuries is that spinal cord neurons don't have the ability to regrow after an injury. That's why most spinal paralysis in patients is permanent.

So scientists tend to focus their research on regrowing peripheral neurons – those that extend from the spinal column to the tips of the hands and feet. Peripheral neurons in the body's extremities have the ability to regenerate, helping people regain some movement and sensation.

In new research, scientists at Washington University School of Medicine in St. Louis have identified a master gene involved in orchestrating the regrowth of peripheral nerves. The gene works as a main switch, making other genes "flip on" in a domino-like fashion. Understanding how these nerves regenerate one day may aid efforts to regrow [spinal cord](#) neurons, the researchers said.

The findings are published online Oct. 29 in the journal *Neuron*.

Surprisingly, senior author Valeria Cavalli, PhD, associate professor of neurobiology, has shown that injury to peripheral nerves flips on a master gene, called hypoxia-inducible factor 1-alpha, otherwise known as HIF-1alpha. This gene, in turn, activates some 200 genes involved in the regrowth of [peripheral nerves](#).

"What is interesting about HIF-1alpha is that it is known to be sensitive to how much oxygen is in the cell," said Cavalli. "If the [oxygen level](#) is normal, there isn't much HIF-1alpha, but in stress conditions such as a lack of oxygen, there is more of this gene to protect the cell and organism from this stress. This knowledge gave us the opportunity to use a noninvasive tool to increase the levels of this factor by simply decreasing the levels of oxygen."

In the lab, Cavalli cultured neurons in a plastic dish and used a zero oxygen environment to measure the regrowth of nerve axons, the branches of nerve cells that transmit nerve signals. In other experiments, the researchers stressed out mice, putting them in 8 percent oxygen

environments for 10 minutes, followed by normal air – about 21 percent oxygen – for 10 minutes. They repeated this cycle six times.

What they found was that the "stressed out" neurons responded by growing the nerve axons that connect to the muscle (motor neurons) as well as those that connect to the skin (sensory neurons) much better than those that were not "stressed out" with low oxygen.

"If you remove the oxygen for a little time, the cell thinks 'I'm so stressed out, I have to do something!' and starts activating many genes that allow the nerves to start to regrow," she said.

Doctors already used a similar technique in a study of patients with chronic, incomplete spinal cord injury—patients whose spinal cords are injured but in such a way that some communication between the brain and spinal cord exists, allowing limited feeling and movement. In that study, doctors encouraged recovery of movement by alternating between a 9 percent oxygen environment for 90 seconds and a typical 21 percent oxygen level for 60 seconds, for 15 cycles. The idea was to "stress" the neurons spared by injury and see if that improved their function.

The hope for Cavalli's team is that the low oxygen environment also may stimulate the damaged neurons to start regenerating and improve recovery.

Cavalli only has seen the nerve axon growth under the microscope. She hopes to keep working to find the right balance and answer the questions: Is less oxygen truly beneficial? What about the time limits? Does functional recovery occur faster and better in mice?

"What we hope, and this is part of our future studies, is that the hypoxia regimen also could wake up the damaged neurons in the spinal cord to start regrowing their axons," she said.

It was surprising to find the HIF-1alpha at the head of the domino chain, Cavalli said.

"It's exciting to find something you can use to manipulate neural growth that's easy and noninvasive like controlling the [oxygen](#) environment," she said. "It could be something we could use to treat nerve injuries and try to get the [neurons](#) to start regenerating."

More information: Activating Injury-Responsive Genes with Hypoxia Enhances Axon Regeneration through Neuronal HIF-1 α . *Neuron*, DOI: [dx.doi.org/10.1016/j.neuron.2015.09.050](https://doi.org/10.1016/j.neuron.2015.09.050)

Provided by Washington University School of Medicine in St. Louis

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