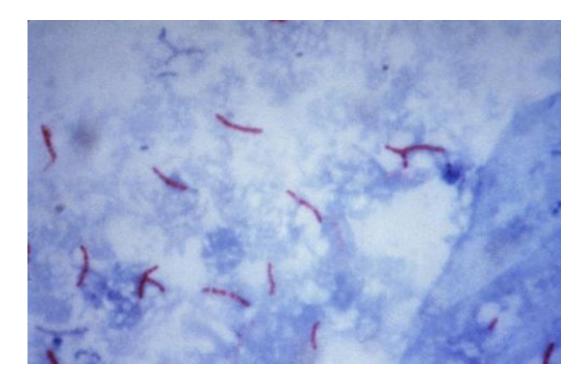


## **Researchers advance treatment of tuberculosis by targeting new enzyme**

November 10 2016



This photomicrograph reveals Mycobacterium tuberculosis bacteria using acidfast Ziehl-Neelsen stain; Magnified 1000 X. The acid-fast stains depend on the ability of mycobacteria to retain dye when treated with mineral acid or an acidalcohol solution such as the Ziehl-Neelsen, or the Kinyoun stains that are carbolfuchsin methods specific for M. tuberculosis. Credit: public domain

Researchers at Johns Hopkins report they have laid the foundation to develop novel antibiotics that work against incurable, antibiotic-resistant bacteria like tuberculosis by targeting an enzyme essential to the



production and integrity of bacterial cell walls.

The findings, they say, suggest that antibiotic drugs specifically targeting the recently discovered *LD-transpeptidase* enzyme, which is needed to build <u>bacterial cell walls</u> in some bacteria, could potentially cure many antibiotic-resistant infections.

An additional implication of the research, the Johns Hopkins team says, is that drugs targeting the enzyme could offer quicker, cheaper and more easily accessible treatment against tuberculosis, a disease that still kills more people worldwide than any other infection, according to the Centers for Disease Control and Prevention.

A summary of the findings is published on Nov. 7 in *Nature Chemical Biology*.

"The complexity of TB treatment and growing prevalence of antibiotic resistance is a serious threat to public health," says Gyanu Lamichhane, Ph.D. associate professor of medicine at the Johns Hopkins University School of Medicine. His team joined with the research team of Craig Townsend, Ph.D., the Alsoph H. Corwin Professor of Chemistry at The Johns Hopkins University's Krieger School of Arts and Sciences, to tackle this complex issue. "Our research offers steps toward the design of new antibiotics that attack a previously untargeted bacterial enzyme," says Townsend.

At the root of their investigation, Lamichhane says, is the fact than more than half of antibiotics prescribed today are of a class called betalactams, which work by interrupting the function of the *DDtranspeptidase* enzyme that creates bacterial cell walls. Without it, bacteria quickly die. However, in 2005, a team of researchers found a second wall-building enzyme, LD-transpeptidase, that allows bacteria like the ones that cause TB to survive <u>antibiotic treatments</u>. "We looked



at the structure of LD-transpeptidase, thought about how it works and started making new compounds that could be used against it," says Townsend.

Pankaj Kumar, Ph.D., postdoctoral fellow in infectious diseases at the Johns Hopkins University School of Medicine, began the research in the new study by extracting LD-transpeptidase from many species of bacteria and examining its detailed molecular structure with a sophisticated imaging system known as protein X-ray crystallography using the Advanced Photon Source at the Argonne National Laboratory in Chicago.

By analyzing the enzyme's structure, Johns Hopkins researchers were able to design new compounds in the carbapenem group, a subclass of the beta-lactam antibiotics that bind to the LD-transpeptidase wallbuilding enzyme and stop its function.

In live bacterial cultures, the carbapenems were shown by Lamichhane's and Townsend's groups to stop the enzyme's wall-building activity. The new compounds were even effective against the ESKAPE pathogens, a group of six bacterial species that the Centers for Disease Control and Prevention has identified as a threat because of their propensity for developing antibiotic resistance.

Following these successes, Amit Kaushik, Ph.D., a postdoctoral fellow in infectious diseases at the Johns Hopkins University School of Medicine, tested two carbapenems in vivo against TB in mice infected with TB.

Researchers infected mice with tuberculosis bacteria and separated them into different treatment groups. The rodents' lungs were sampled periodically over a period of three weeks, and the results showed that even without use of classic TB antibiotic treatments, the new carbapenems, specifically biapenem, cured TB infection in mice.



"Our data show that the carbapenems successfully treated TB infections alone by attacking the right enzyme," says Lamichhane.

Townsend and Lamichhane say the focus of their research is now on creating variations of their original compound that are designed to target specific bacteria. Many commonly prescribed antibiotics today work on a broad spectrum of bacterial species, meaning that in addition to killing off bad bacteria, they also destroy the friendly bacteria our bodies need to function normally and whose destruction can cause dangerous side effects. Lamichhane believes that the future of antibiotic treatments relies on our ability to be good antimicrobial stewards and treat specific bacteria without affecting our bodies' natural microbiome. Not only will this cut down on antibiotic resistance in the bacterial species not being targeted.

The researchers are now in the process of initiating clinical trials to test the safety and efficacy of some of these new compounds.

Antibiotic resistance has been an ever-present threat since the discovery of penicillin in 1928. Scientists and physicians have historically kept up with resistant bacteria through the frequent introduction of new antibiotic treatments. However, the research and development of antibiotics <u>plummeted in the 1980s</u> as other drugs became more profitable to produce. Because of the decreased incentive to invest in new antibiotics and the <u>liberal use of antibiotics</u> we already have, many <u>bacterial species</u> have quickly outpaced our ability to treat them. The Centers for Disease Control and Prevention estimates that annually, 2 million illnesses and 23,000 deaths are caused by <u>antibiotic-resistant</u> <u>bacteria</u> in the United States alone, and that 70 percent of hospital-acquired infections are now antibiotic-resistant.

More information: Pankaj Kumar et al. Non-classical transpeptidases



yield insight into new antibacterials, *Nature Chemical Biology* (2016). DOI: 10.1038/nchembio.2237

## Provided by Johns Hopkins University School of Medicine

Citation: Researchers advance treatment of tuberculosis by targeting new enzyme (2016, November 10) retrieved 11 February 2023 from https://medicalxpress.com/news/2016-11-advance-treatment-tuberculosis-enzyme.html

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